Current Chemistry Letters 11 (2022) 147-156

Contents lists available at GrowingScience

Current Chemistry Letters

homepage: www.GrowingScience.com

Improving seed germination and seedling growth of maize (Zea mays, L.) seed by soaking in water and moringa oleifera leaf extract

Abeer A. Ahmed^{a*} and Amal A. El-Mahdy^a

^aSeed Technology Research Department, Field Crops Res. Inst., Agriculture. Res. Center (ARC), Giza. Egypt

CHRONICLE	A B S T R A C T
Article history: Received August 22, 2021 Received in revised form October 25, 2021 Accepted February 22, 2022 Available online February 22, 2022 Keywords: Zea mays L. Germination Seedling growth Soaking duration Moringa oleifera leaf extract	Germination traits are the key factor in higher productivity of crops. Thus, laboratory tests were made to enhance the germination of the seeds. One of the best strategies to promote seed germination was soaking the seeds in water and Moringa leaves extracts (MLE.) before planting. This experiment was carried out to evaluate the optimal soaking duration to enhance seed germination and seedling growth of hybrid 178 maize. There were three soaking durations; 12 hours, 16 hours, and 20 hours, along with the un-treatment (0 hour or without soaking). While the other factor was the concentration of the soaking seeds solution in Moringa leaves extract which was (0.25, 0.5 and 1.00 g) at the ratio of 1:10 (w/v) beside to hydro-priming (soaking in water) and control (un-treatment), each with four replications. Seed germination (%), germination index (GI), germination speed index (SGI), germination rate (GR, day), mean germination rate (MGR, day), seedling shoot length (cm), seedling vigor 1 (SV1) and seedling vigor 2 (SV2) were evaluated. Results showed that maximum value germination traits were found in hybrid 178 of Maize which soaked at 20 hours of duration, followed by soaked at 16 hours then at 12 hours soaking in Moringa leaves extract. 0.25 g/mm MLE. was the best treatment, followed by 0.5 (g) and hydro-priming gave higher values, as to seed germination and seedling vigour (SVII) with 0.25 g/mm then 0.5 g/mm and hydro-priming under soaked seed to 20 hour com. This suggests that soaking in Moringa leaves extract as 0.25 concentrate under 20 hour soaking was a suitable treatment that can improve germination and seedling growth in hybrid 178 maize.

© 2022 by the authors; licensee Growing Science, Canada.

1. Introduction

Different applications for organic compounds in agriculture and other fields are well-known.¹⁻¹⁸ Seed priming is a presoaking treatment which aids in the physiological process that allows seeds to germinate more efficiently. Priming techniques such as hydro-priming, osmo-priming, chemical priming, hormonal priming, and biological priming have all been used. Seed priming is a low-cost technique that involves soaking the seeds in a solution for a specific amount of time after which they are hydrated to allow the metabolic process of germination to occur while preventing the emergence of radicals. The seeds perform well under both normal and stressful conditions.

^{*} Corresponding author. E-mail address: <u>abeerabdelaty2015@gmail.com</u> (A. A. Ahmed)

^{© 2022} by the authors; licensee Growing Science, Canada doi: 10.5267/j.ccl.2022.2.005

Germination stage is a direct indicator of higher crop production. The main focus for increased maize productivity is to promote maize germination. ³⁹ reported that prolonged germination duration could result in late emergence of seedlings as a result of direct interaction with soil-borne pathogenic bacteria. The amount of oxygen and light supplied by the soil, as well as other edaphic factors. Most farmers believe that pre-soaking seed in water before planting improves germination.²⁸ Soaking seeds in water aims to reduce the lag-phase in seedling germination, which can harm seedlings and result in a noticeable decrease in crop productivity.

Soaking seed prior to planting reduced the negative effects of high temperatures and improved seed germination. Furthermore, it provides a sufficient amount of moisture content to the seed, which may improve germination rate.³⁰ Some tree extracts and crop residues have been shown to influence crop growth and yield.^{29,22, 26} M. oleifera leaf extract has been shown to accelerate plant growth in the early stages, strengthen plants, improve resistance to diseases and pests, increase leaf area duration, produce larger and more fruits, and increase overall yield productivity by 20 to 35 percent.³¹ With high global prices for inorganic fertiliser, water pollution and land degradation associated with inorganic fertiliser use, and the contribution of climate change, there is a need to search for alternative plant nutrient sources. Moringa is one of the important alternatives being studied to determine its effect on crop growth and productivity, so that M. oleifera can be improved as a possible supplement or substitute for inorganic fertiliser, and it has begun to be promoted as a multipurpose plant among farmers.⁴⁵ Moringa leaf extract was sprayed on the leaves of bell pepper, soyabeans, onions, sorghum, tea, melon, maize, and chilli, and it increased crop yield.³¹ MLE application has proven to be an extremely valuable source of plant growth-promoting substances. Moringa oleifera extracts are either used as a seed priming agent or as a foliar spray to promote growth.^{40, 43}

The purpose of this study was to compare the effect of soaking in M. oleifera leaf extracts on germination traits and early seedling growth of hybrid 178 maize in Egypt to soaking in water and untreated seed (control).

2. Material and Methods

Experiments of laboratory and field emergence were conducted at the Seed Tech. Res. Laboratory, Seed Technology Research Department, Field Crops Research Institute, Agriculture Research Center, and at Greenhouse of Agriculture Research Center, Giza, Egypt. The study aims to evaluate the effect of seed soaking durations (control, 12, 16 and 20 hours) under different concentration (0.25, 0.50 and 1.00 g at the ratio of 1:10 (w/v) from Moringa leaves extract) beside to soaking in water only (hydro-priming) and control (untreated seed) on germination parameters, seedling vigor and field emergence of maize (*Zea maize*, L.). The experiment was laid out in completely randomized design (CRD) with four replications. Maize seeds (hybrid 178) were obtained from Maize Research Department, FCRI, ARC. Experiment was artificially created in the sterilized Petri dishes. Four replicates of 20x5 seeds each from every treatment were planted in 15-cm diameter Petri dishes moistened with distilled water, incubated in a growth chamber at 25°C and laid in factorial Completely Randomized Design (CRD). Normal seedlings were counted and then germination percentage was calculated according to the rules outlined before.³⁵

2.1 Preparation of MLE

Fresh M. oleifera leaves were air-dried before being processed into powder. The crude powders were kept at room temperature in paper bags. Moringa leaves powder was soaked in distilled water for 24 hours at room temperature $(20 \pm 2^{\circ}C)$ with intermittent shaking to obtain stock moringa leaves extract (0.25, 0.5, and 1 g at a ratio of 1:10 (w/v) turn to 100mm). To eliminate fibre debris, the mixture was filtered through four layers of cheesecloth, followed by Whatman No.1 filter paper. In addition to the control (distilled water), three different concentrations of MLE (i.e., 0.25, 0.5, and 1 g at a ratio of 1:10 (w/v) turn to 100mm) were created. The mineral content and chemical composition in Moringa leaf extract were summarized in Table (1 and 2).

Mineral contents (mg/100g.d.wt)								
Essential macro-elements				Esse	ntial micro-eler	nents		
N (g/100g)	Р	k	Mg	Ca	Fe	Cu	Zn	
1.78	9.7	2.8	3.5	1.28	1.18	0.87	2.46	

Table 1 . The mineral contents of Moringa Leaf Extract (M	ALE).	33
--	-------	----

Table 2. Chemical composition analysis of moringa leaf extract. ¹⁹

Chemical composition	(mg/100g. d.wt)	Chemical composition	(mg/100g. d.wt)
Water	5.90	Ascorbic acid	3.26
Protein	27.20	Total carotenoids	2.24
Fiber	19.20	Soluble phenols	2.24
Total sugar	38.60	Gibberellins	0.802
Lipids	17.10	Zeatin	0.936

2.2 Seed preparation

To remove any traces, seeds were disinfected for 5 minutes with a 0.1 percent HgCl2 solution before being rinsed 5-6 times with distilled water. This was achieved by randomly selecting 20 viable seeds from each replicate and soaking them in each MLE treatment solution and hydro-priming them for 12, 16, and 20 hours before rinsing them with distilled water and air drying them for 24 hours before planting them in the laboratory.

Four replications of 100 seeds (each replicate 5 times x20 seeds) of treated and untreated maize seeds were sown in each sub replication in sterilised Petri dishes covered at the bottom with two sheets of Whitman filter paper, then placed in an incubator at $25\pm2^{\circ}$ C for 7 days, according to ISTA rules.³⁹ The total number of seeds that germinated was tallied every day, and the percentage was computed on the seventh day. Germination (%): (The number of germinated seeds/The total number) × 100.

Germination speed test: For each replicate, seeds were inspected daily and considered germinated following radical emergence. The seeds that had germinated were counted and removed from the Petri dishes. Speed Germination Index (SGI), it was calculated as described in the Association of Official Seed Analysis²³ by following formula:

SGI = (No. of germinated seed/days of first count) + (....) + (No. of germinated seed/days of final count). When the radical of a seed was at least 2 mm long, it was deemed germinated.

Germination Rate (GR), it was defined according to the following formula:

 $GR = a + (a + b) + (a + b + c) \dots (a + b + c + m)/n (a + b + c + m)$. Where a, b, c are No. of seedlings in the first, second and third count, m is no. of seedlings in final count, n is the number of counts.²⁵

Mean Germination Time (MGT), it was calculated based on the following equation: $MGT = \Sigma Dn / \Sigma n$, where (n) is the number of seeds which were germinated on day (D) is number of days counted from the beginning of germination.²⁷

Ten normal seedlings were used to measure seedling root and shoot length (cm) at 7 days after soaking. While ten typical seedlings were measured for fresh and dry weight (g) 7 days after planting, the seedlings were dried in a hot-air oven at 85° C for 12 hours to determine the seedlings dry weight (g).

The product of the germination percentage, seedling length, and seedling dry weight was used to calculate seedling vigour. Seedling vigour was estimated as follows:

Vigour index I = Germination (%) x Seedling length (Root + Shoot).

Vigour index II = Germination (%) x Seedling dry weight (Root + Shoot).²⁰

2.3 Field emergence experiment

Field emergence (percent), seedling dry weight, and seedling vigour after 14 days from planting were all tested characters in a screen house experiment.

2.4 Statistical analysis

Statistical analysis of the results was performed using analysis of variance ANOVA and least significant differences (L.S.D.) were obtained using ANOVA tables.⁴⁸

3. Results

Soaking of seeds in different times (12, 16 and 20 hour), resulted in a significant effect on the germination traits compared to- non soaking seed as shown in **Table (3)**. The seed germination increased to 14.2, 13.6 and 13.2 % for 20, 16 and 12 h receptively, compared with non-soaked seed. Soaking at 20 h was the best compared with other treatments. The results indicated that Germination Rate (GR; day) was significant and increased to 11.5, 7.7 and 5.7 % by 20, 16 and 12 h soaking, respectively. While, it was decreased the mean germination time with soaking treatments to 3.0 for 20 h, 3.2 for 16 h and 3.2 for 12 h compared with control which was 3.3. However, the speed germination index was significant between all treatments; the higher speed germination was increased by 20.1 % to 20 h then 17.4 to 16 h then 15.0 to 12 h.

Table 3. Effect of different soaking durations on the germination traits of maize seeds.

Soaking duration	GP (%)	Increase% over	GR/day	Increase% over	MGT/ day	Increase % over	SGI	Increase % over
		control		control	·	control		control
12 hour	80.3 b	13.2	0.55 b	5.8	3.2 a	-5.3	15.3 b	15.0
16 hour	80.5 ab	13.5	0.56 b	7.7	3.2 a	-5.6	15.7 ab	17.4
20 hour	81.0 a	14.2	0.58 a	11.5	3.0 b	-7.3	16.0 a	20.1
Control	70.9 c		0.52 c	_	3.3 a	-	13.3 c	_
F test	**		**		**		**	
LSD 5%	0.56		0.013		0.06		1.77	

GP: Germination Percentage, GR: Germination Rate, MGT: Mean Germination Time, SGI: Speed Germination Index.

Soaking duration significantly affected the seedling characters of hybrid 178 maize. Seedling characters (shoot, root and seedling length) were recorded lower values with unsoaked seeds while, it gave higher value with seed soaking at 20 h ,which were increased to 21.0, 19.2 and 22.7% for shoot, root and seedling length respectively, comparing with 16 h (16.3% for shoot, 15.0% for root and 18.0% for seedling), and 12 h soaking which was (13.4% for shoot, 9.0% for root and 13.9% for seedling) (Table 4).

Data in **Table (5)** showed that soaking treatments had an effect on seedling fresh and dry weight. However, heavier fresh seedling was achieved by soaking 24.0, 20.8 and 12.5% for 20, 16 and 12 h respectively, whereas minimum fresh weight was recorded for non-soaked seeds. The same trend with dry weight which recorded higher values 25.0, 20.0 and 15.0% with 20, 16 and 12 h respectively compared with control.

Maximum seedling vigor increase was obtained in seed soaking for 20 h, which was 22.8% and 17.0% for SV1 and SV2 respectively. Followed by 16 h was increased to 11.7% and 14.8% for SV1 and SV2 respectively then 12 h was 6.2% and 12.1% with SV1 and SV2 respectively compared with control (non-soaked) (Table 5).

Soaking	Seedling length (cm)								
duration	Shoot length	Increase% over control	Root length	Increase% over control	Seedling length	Increase % over control			
12 hour	12.5 a	13.4	7.1 ab	9.0	19.6 b	13.9			
16 hour	12.8 a	16.3	7.5 a	15.0	20.3 a	18.0			
20 hour	13.3 a	21.0	7.8 a	19.2	21.1 a	22.7			
Control	11.0 b		6.5b		17.2 c				
F test	**		**		**				
LSD 5%	1.00		0.72		1.21				

Table 4. Effect of different soaking durations on the Seedling length (cm) of maize seeds.

Seedling weight (g)					weight (g) Seedling vigor					
Soaking duration	Fresh	Increase% over control	Dry	Increase% over control	SV1	Increase % over control	SV2	Increase % over control		
12 hour	1.08ab	12.5	0.23 a	15.0	1637.0 bc	6.2	20.4 ab	12.1		
16 hour	1.16 a	20.8	0.24 a	20.0	1721.6 ab	11.7	20.9 a	14.8		
20 hour	1.19 a	24.0	0.25 a	25.0	1891.6 a	22.8	21.3 a	17.0		
Control	0.96 b		0.20 b	_	1541.1 c	_	18.2 b	_		

Table 5. Effect of different soaking durations on the Seedling weight (g) and Seedling vigor of maize seeds.

Using MLE treatments were effective in increasing all germination and seedling traits. Germination % in Table 6 was recorded large value with using 0.25g and 0.50g concentrate reach to 28.12 and 28.08 % increasing then hydro-priming and 1g which were 14.66 and 0.31 % respectively, compared with control (untreated seed).

**

171.7

**

2.58

Data showed that 0.25 was significantly greater than 0.50, hydro-priming and 1 g which were 5.6, 3.7, 3.7 and 1.8 respectively, compared with control for germination rate trait.

Mean germination time was recorded with high values with control and smallest values with 0.25 g which was -3.9 %. while increasing speed germination seed as compared to that of untreated seeds and gave high speed germination with 0.25g (34. 9% increase) and lowest speed with 1 g (11.2 % increase) compared with control (Table 6).

**

0.029

MLE.	GP	Increase%		Increase%		Increase		Increase
Conc.	(%)	over	GR/day	over	MGT/day	% over	SGI	% over
(g/mm)		control		control	MG1/uay	control		control
0.25 g/10mm	90.3 a	28.1	0.57 a	5.6	3.17 b	-3.9	17.6 a	34.9
0.50 g/10mm	90.2 a	28.0	0.56 ab	3.7	3.17 b	-3.9	16.8 ab	28.6
1.00 g/10mm	70.7c	00.3	0.55 bc	1.8	3.19 b	-3.3	14.5 bc	11.2
Water	80.7 b	14.7	0.56 ab	3.7	3.18 b	-3.6	16.5 ab	26.3
Control	70.4 c	_	0.54 c	_	3.30 a	_	13.0 c	_
F test	**		**		**		**	
LSD 5%	0.72		0.017		0.09		2.28	

MLE.Conc.: Moringa Leaves Extract Concentration, GP: Germination percentage, GR: Germination Rate, MGT: Mean Germination Time, SGI: Speed Germination Index.

F test

LSD 5%

**

0.19

151

MLE.	Seedling length (cm)					
Conc. (g/mm)	Shoot	Increase% over control	Root	Increase% over control	Seedling	Increase % over control
0.25 g/10mm	13.6 a	17.2	8.4a	14.4	21.9a	11.1
0.50 g/10mm	13.5 a	16.3	7.6b	4.7	21.0a	6.3
1.00 g/10mm	12.3b	5.6	7.3d	00.0	19.7b	0.00
Water	13.2a	13.0	7.4c	1.6	20.8a	5.5
Control	11.6c		7.3d	-	19.7b	-
F test	**		**		**	
LSD 5%	0.85		0.09		1.00	

Table 7. Effect of Moringa oleifera leaf extract (MLE) on the Seedling length (cm) of maize seeds.

MLE.Conc.: Moringa Leaves Extract Concentration.

Table 8. Effect of Moringa	<i>leifera</i> leaf extract	(MLE) on the Seedling	g weight (g) and See	dling vigor of maize seeds.
----------------------------	-----------------------------	-----------------------	----------------------	-----------------------------

MLE.	Seedling weight (g)				Seedling vigor			
Conc. (g/mm)	Fresh	Increase % over control	Dry	Increase % over control	SV1	Increase % over control	SV2	Increase % over control
0.25 g/10mm	0.119 a	21.7	0.027 a	22.7	2091.9 a	48.9	22.6 a	30.1
0.50 g/10mm	0.119 a	21.7	0.024 ab	9.1	1940.1 a	38.1	21.5 a	23.5
1.00 g/10mm	0.113 c	11.3	0.022 b	0.0	1479.0 c	5.30	20.7 a	19.3
Water	0.114 b	16.3	0.023 ab	4.6	1834.6 b	30.6	21.4 a	23.0
Control	0.098 d	_	0.022 b	_	1404.5 c	_	17.4 b	_
	**		*		**		**	
LSD 5%	0.035		0.04		221.69		2.33	
MLE Cana - N	f	Entra to						

MLE.Conc.: Moringa Leaves Extract Concentration.

Most seed treatments resulted in higher seedling fresh weight compared with that of control whereas seedling dry weight was a non-significant increase with 1 g by priming compared with control (Table 8).

Maximum seedling vigour (SVI and SVII) was improved with all treated seed, which was 48.9, 38.1, 30.6 and 5.3% Increase over control by 0.25, 0.5, hydro-priming and 1 g respectively, for SV1. Whereas, it was increased by 30.1, 23.5, 23.0 and 19.3% Increase over control for 0.25, 0.5, hydro-priming and 1g for SV2 compared with control (**Table 8**).

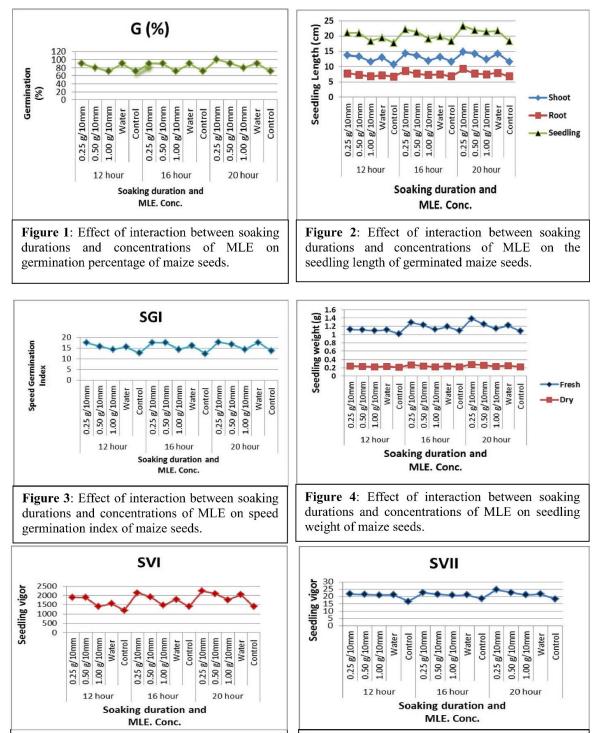
Data indicated that significant interaction between soaking duration and MLE for all germination traits and seedling growth.

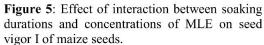
Significant differences were shown in recorded germination % trait with interaction between soaking duration and MLE treatments.0.25 g MLE under 20 h soaked gave the highest value of all recorded parameters (100.0), followed by 0.5 g MLE and hydro-priming compared with other treatments and control. Whereas the lowest value was 70.67 for untreated seed in **Fig. (1)**. The present study also indicates that all the treated seed significantly improved shoot, root and seedling length, however, treated with 0.25 g MLE success to improve these traits and gave higher values under 20 h (14.3, 9.2 and 22.2 for shoot, root seedling respectively.) and 16 h which were (14.8, 8.5 and 23.3 for shoot, root seedling respectively.) and low values were for untreated seed (**Fig. 2**).

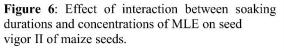
The results showed that increasing speed germination seed as compared to that of untreated seeds (Fig. 3) and gave high speed germination with 0.25g MLE (17.9) under 20h and lowest speed with 1 g MLE followed by control which was the lowest speed.

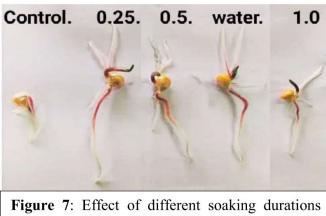
Most of seed treatments resulted in higher seedling fresh and dry weight compared with control, seedling dry weight was non-significant with 0.25 g MLE under 20 and 16 hour soaking followed by 0.5g and hydro-priming comparing whereas, untreated seed had low number comparing with other treatments and control (Fig. 4).

Maximum seedling vigor was seen in seed soaking for 20 h with 0.25 g (2245.2 and 23.6 for SVI and SVII (20 h) respectively and 2245.2 and 22.6 for SVI and SVII, (16 h), respectively) then 0.5 and hydro-priming under 20 h compared with non-soaked (Fig. 5 and Fig. 6).









and MLE concentrations on seedling traits after three days from germination of seed maize.

Field emergence experiment

Table 9 shows significant effects of soaking duration, MLE. Conc. (g/mm) and its interaction treatments on hybrid 178 maize field emergence.

Table 9. Effect of seed soaking duration and <i>Moringa oleifera</i> leaf extract (MLE) on field emergence (%), seedling dry
weight (g pot ⁻¹) and seedling vigor (SVII) parameters of hybrid 178 maize.
Treatment

	Treatment			
	MLE.	Field	Seedling dry wt.	Seedling
Soaking duration	Conc.	emergence (%)	(g)	vigor (SVII)
	(g/mm)			
12 hour	0.25 g/mm	10	0.21	21.7
	0.50 g/mm	10	0.21	20.2
	1.00 g/mm	9	0.20	21.0
	Water	9.5	0.21	20.1
	Control	9.5	0.19	19.1
16 hour	0.25 g/mm	9.5	0.25	24.6
	0.50 g/mm	10	0.24	22.2
	1.00 g/mm	9.5	0.22	21.7
16	Water	10	0.23	21.8
	Control	9	0.22	21.5
	0.25 g/mm	9.5	0.27	26.2
20 hour	0.50 g/mm	10	0.26	25.6
ho	1.00 g/mm	9.5	0.24	21.7
20	Water	9.5	0.25	25.0
	Control	9.5	0.23	19.7
F test		ns	**	**
LSD 5%		1.25	0.03	2.73
	12 hour	9.6	0.21	20.3
Soaking hour	16 hour	9.6	0.23	22.4
	20 hour	9.6	0.24	22.8
F test		ns	**	**
LSD 5%		0.56	0.016	1.32
	0.25 g/mm	10	0.25	22.9
	0.50 g/mm	9.6	0.23	22.0
Concentration	1.00 g/mm	9.5	0.22	20.7
	Water	9.5	0.23	21.9
	Control	9.3	0.19	19.7
F test		ns	**	**
LSD 5%		0.72	0.01	0.73

Regarding germination % in **Table 9**, there was non-significant with soaking and MLE. Conc. (g/mm) and interaction between them. While Seedling dry wt. gave significant increases which were obtained through soaked seeds 20 h (0.24 g) followed by 16 h (0.23 g). Whereas it was increased with 0.25 g MLE (0.25g) followed by 0.5 g MLE (0.23g) and hydropriming (0.23g) compared to 12 hour soaked, 0.1 g MLE and control treatments in barley field emergence. Addition to the best dry weight in interaction was between soaked in 0.25 g MLE under 20 hours (0.27g). The increment reached to 22.8

and 22.4% by soaked 20 h and 16 h and retracted to 22.9 by 0.25 MLE followed by 22.0 and 21.9 by 0.5g /mm and hydropriming, respectively in field emergence for Seedling vigor (SVII). Significant differences were showed in recorded maize traits at field emergence with interaction between soaking duration and MLE treatments.0.25 g MLE under 20 h soaked gave the highest value of all recorded parameters (25.2), followed by 0.5 g MLE (25.6) and hydro-priming (25.0) compared with other treatments and control in **Table 9**.

4. Discussion

Rate of germination, germination percentage, speed germination and seedling biomass are important contributors of seed vigour. Increasing emergence rate is the main foundation, which ensures improving of overall seedling performance. Our results showed that optimum soaking time (20 then 16 hours) was better than 12 h and control using MLE concentrate (0.25 g/mm). According to⁵⁰ most seeds treated in water before sowing resulted in faster germination. Additionally,⁴⁴ showed that sufficient moisture is required for quick seed germination. Higher germination (percentage) was also mentioned as an important factor in determining the amount and viability of seeds.⁴² This mechanism was triggered by high water availability, which induced leaching of important soluble food reserves in the seeds, as well as exosmosis of hormones and enzymes, which lowered respiration and protein synthesis rates.³⁷

Hydrolysis of complex carbohydrates into simple sugars, which are easily consumed during protein synthesis and auxins, increases germination. The development of new tissues is aided by the production of additional auxins, which softens the cell wall and supports protein use and growth. These results are ascribed to moringa leaf extract (MLE), which contains plant hormones that cause seed germination to improve.⁴⁷ Furthermore,^{34,46} ensure that moringa leaf extract (MLE) is a source of vital amino acids as well as various minerals such as N, K, Zn, and Ca, all of which are considered natural growth boosters. Moringa leaf extract was tested for mineral content and chemical composition according to^{33, 19} who reported that moringa leaves content on essential minerals as P, K, Mg, Ca, Fe, Cu and Zn in addition to ascorbic acid, lipids, soluble phenols, gibberellins and zeatin. In contrast to the 2 percent and 4 percent values, the high level of MLE (6%) has an inhibitory influence on prior parameters.³³ According to³⁸ soaking maize seeds in a high concentration of MLE solution caused imbibition damage, which resulted in a decrease in germination features. These results are attributable to the seed being treated with MLE solution, which causes a variety of biochemical changes such as starch hydrolysis, enzyme activation, and dormancy break.²⁴ MLE extract, on the other hand, improves the mobilisation of reserves from seed storage as endosperms or cotyledons for partitioning to embryo by lowering sugars and increasing amylase activity, adding to early seed vigour.²¹ According to reports, the MLE extract can boost germination percentage, index, and rate.^{49,41} According to⁴¹ the effect of activity, MLE solution, can be linked to the presence of growth-promoting chemicals in MLE extract, which increased seed germination features and seedling growth. This paper can be considered as a good evidence for the importance of scientific research in agriculture and other fields.⁵¹⁻⁵⁸

5. Conclusion

In general, soaking hybrid 178 maize seeds in Moringa leaf extract at a concentration of 0.25 g/mm was the superior treatment for achieving the highest values of germination traits and early seedling growth. Furthermore, by using Moringa leaf extract at a concentration of 0.25 g/mm for 20 /or 16 hour durations as a seed soaking treatment, it can stimulate the rapid of germination, promote germination percentage, and have positive effects on seedlings growth, resulting in healthy and strong seedlings.

References

- Abdel-Raheem Sh. A. A., Kamal El-Dean A. M., Zaki R. M., Hassanien R., El-Sayed M. E. A., Sayed M., and Abd-Ella A. A. (2021) Synthesis and toxicological studies on distyryl-substituted heterocyclic insecticides. *Eur. Chem. Bull.*, 10 (4) 225-229.
- 2 Kamal El-Dean A. M., Abd-Ella A. A., Hassanien R., El-Sayed M. E. A., and Abdel-Raheem Sh. A. A. (2019) Design, Synthesis, Characterization, and Insecticidal Bioefficacy Screening of Some New Pyridine Derivatives. ACS Omega, 4 (5) 8406-8412.
- 3 Tolba M. S., Sayed M., Kamal El-Dean A. M., Hassanien R., Abdel-Raheem Sh. A. A., and Ahmed M. (2021) Design, synthesis and antimicrobial screening of some new thienopyrimidines. *Org. Commun.*, 14 (4) 334-345.
- 4 Abdel-Raheem Sh. A. A., Kamal El-Dean A. M., Hassanien R., El-Sayed M. E. A., and Abd-Ella A. A. (2020) Synthesis and biological activity of 2-((3-Cyano-4,6-distyrylpyridin-2-yl)thio)acetamide and its cyclized form. *Alger. j. biosciences*, 01 (02) 046-050.
- 5 Abdel-Raheem Sh. A. A., Kamal El-Dean A. M., Abdul-Malik M. A., Hassanien R., El-Sayed M. E. A., Abd-Ella A. A., Zawam S. A., and Tolba M. S. (2022) Synthesis of new distyrylpyridine analogues bearing amide substructure as effective insecticidal agents. *Curr. Chem. Lett.*, 11 (1) 23-28.
- 6 Bakhite E. A., Abd-Ella A. A., El-Sayed M. E. A., and Abdel-Raheem Sh. A. A. (2017) Pyridine derivatives as insecticides. Part 2: Synthesis of some piperidinium and morpholinium eyanopyridinethiolates and their insecticidal activity. J. Saud. Chem. Soc., 21 (1) 95–104.
- 7 Kamal El-Dean A. M., Abd-Ella A. A., Hassanien R., El-Sayed M. E. A., Zaki R. M., and Abdel-Raheem Sh. A. A. (2019) Chemical design and toxicity evaluation of new pyrimidothienotetrahydroisoquinolines as potential insecticidal agents. *Toxicol. Rep.*, 6 (2019) 100-104.

A. A. Ahmed and A. A. El-Mahdy / Current Chemistry Letters 11 (2022)

- 8 Gad M. A., Aref S. A., Abdelhamid A. A., Elwassimy M. M., and Abdel-Raheem Sh. A. A. (2021) Biologically active organic compounds as insect growth regulators (IGRs): introduction, mode of action, and some synthetic methods. *Curr. Chem. Lett.*, 10 (4) 393-412.
- 9 Abdel-Raheem Sh. A. A., Kamal El-Dean A. M., Hassanien R., El-Sayed M. E. A., and Abd-Ella A. A. (2021) Synthesis and characterization of some distyryl-derivatives for agricultural uses. *Eur. Chem. Bull.*, 10 (1) 35-38.
- 10 Tolba M. S., Sayed M., Abdel-Raheem Sh. A. A., Gaber T. A., Kamal El-Dean A. M., and Ahmed M. (2021) Synthesis and spectral characterization of some new thiazolopyrimidinederivatives. *Curr. Chem. Lett.*, 10 (4) 471-478.
- 11 Al-Taifi E. A., Abdel-Raheem Sh. A. A., and Bakhite E. A. (2016) Some reactions of 3-cyano-4-(p-methoxyphenyl)-5-oxo-5,6,7,8-tetrahydroquinoline-2(1H)-thione; Synthesis of new tetrahydroquinolines and tetrahydrothieno[2,3-b]quinolines. Assiut University Journal of Chemistry (AUJC), 45 (1) 24-32.
- 12 Abdel-Raheem Sh. A. A., Kamal El-Dean A. M., Hassanien R., El-Sayed M. E. A., Sayed M., and Abd-Ella A. A. (2021) Synthesis and spectral characterization of selective pyridine compounds as bioactive agents. *Curr. Chem. Lett.*, 10 (3) 255-260.
- 13 Abdel-Raheem Sh. A. A., Kamal El-Dean A. M., Abdul-Malik M. A., Abd-Ella A. A., Al-Taifi E. A., Hassanien R., El-Sayed M. E. A., Mohamed S. K., Zawam S. A., and Bakhite E. A. (2021) A concise review on some synthetic routes and applications of pyridine scaffold compounds. *Curr. Chem. Lett.*, 10 (4) 337-362.
- 14 Tolba M. S., Kamal El-Dean A. M., Ahmed M., Hassanien R., Sayed M., Zaki R. M., Mohamed S. K., Zawam S. A., and Abdel-Raheem Sh. A. A. (2022) Synthesis, reactions, and applications of pyrimidine derivatives. *Curr. Chem. Lett.*, 11 (1) 121-138.
- 15 Abdelhafeez I. A., El-Tohamy S. A., Abdul-Malik M. A., Abdel-Raheem Sh. A. A., and El-Dars F. M. S. (2022) A review on green remediation techniques for hydrocarbons and heavy metals contaminated soil. *Curr. Chem. Lett.*, 11 (1) 43-62.
- 16 Tolba M. S., Abdul-Malik M. A., Kamal El-Dean A. M., Geies A. A., Radwan Sh. M., Zaki R. M., Sayed M., Mohamed S. K., and Abdel-Raheem Sh. A. A. (2022) An overview on synthesis and reactions of coumarin based compounds. *Curr. Chem. Lett.*, 11 (1) 29-42.
- 17 Bakhite E. A., Abd-Ella A. A., El-Sayed M. E. A., and Abdel-Raheem Sh. A. A. (2014) Pyridine derivatives as insecticides. Part 1: Synthesis and toxicity of some pyridine derivatives against Cowpea Aphid, Aphis craccivora Koch (Homoptera: Aphididae). J. Agric. Food Chem., 62 (41) 9982–9986.
- 18 Abdelhamid A. A., Elsaghier A. M. M., Aref S. A., Gad M. A., Ahmed N. A., and Abdel-Raheem Sh. A. A. (2021) Preparation and biological activity evaluation of some benzoylthiourea and benzoylurea compounds. *Curr. Chem. Lett.*, 10 (4) 371-376.
- 19 Abdalla M. M. (2014) Boosting the growth of rocket plants in response to the application of as a bio stimulant. *Life Sci. J.*, 11 (11) 1113-1121. Moringa oleifera extracts
- 20 Abdul-Baki A. A., and Anderson J. O. (1973) Vigour determination of soybean seed by multiple criteria. Crop Sci., 13 630-633.
- 21 Afzal I., Hussain B., Basra S. M., and Rehman H. (2012) Priming with MLE reduces imbibitional chilling injury in spring maize. Seed Sci. and Technol., 40 (2) 271-276.
- 22 Ahmed D. M., and Nimer A. M. (2002) Effects of Acacia Senegal (L., Wild) on sandy soils. A case study of El Damokya Forest, Northern Kordofan State. *Univ. Khartoum J. Agric. Sci.*, 10 106-118.
- 23 AOSA. (1983) Association of Official Seed Analysts. Seedvigour testing hand book. Contribution, No. 32 to the Handbood of seed testing.
- 24 Aziza A., Haben A., and Becker M. (2004) Seed priming enhances germination and seedling growth of barley under condition of P and Zn deficiency. J. Plant Nutr. Soil Sci., 167 630-636.
- 25 Bartlett M. S. (1937) Some examples of statistical methods of research in agriculture and applied biology. *Suppl. J. R. Stat. Soc.*, 4 (2) 137-183.
- 26 El Atta H. A., and Bashir I. A. (1999) Adverse effects of Eucalyptus camaldulensis (Dehn) leaf extract on germination and seedling growth of wheat. J. Agric. Sci., 7 (2) 70-80.
- 27 Ellis R. H., and Roberts E. H. (1981) The quantification of ageing and survival in orthodox seeds. Seed Sci. Technol., 2 373-409.
- 28 Esmaeilpour A., and Damme P. V. (2016) Evaluation of seed soaking times on germination percentage, germination rate and growth characteristics of pistachio seedlings. *Acta Hortic.*, 1109 107-112.
- 29 Farooq M., Jabran K., Rehman H., and Hussain M. (2008) Allelopathic effects of rice on seedling development in wheat, oat, barley and berseem. *Allelopath. J.*, 22 385-390.
- 30 Finch-Savage W. E., Dent K. C., and Clark L. J. (2004) Soak conditions and temperature following sowing influence the response of maize (Zea mays L.) seeds to on-farm priming (pre-sowing seed soak). Field Crops Res., 90 (2/3) 361-374.
- 31 Fuglie L. J. (2000) New Uses of Moringa Studied in Nica ragua: ECHO's Technical Network Site-networking global hunger solutions. ECHO. Development Notes (EDN), 1-6. <u>http://edn.link/edn-68</u>
- 32 Fuglie L. J. (1999) The Miracle Tree: *Moringa oleifera*: Natural Nutrition for the Tropics. Revised edition. Church World Service, Dakar. pp. 68.
- 33 Abou El-Nour H. H., and Ewais N. A. (2017) Effect of Moringa oleifera Leaf Extract (MLE) on Pepper Seed Germination, Seedlings Improvement, Growth, Fruit Yield and its Quality. Middle East J. Agric. Res., 6 (2) 448-463.
- 34 Howladar S. M. (2014) A novel Moringa oleifera leaf extract can mitigate the stress effects of salinity and cadmium in bean (*Phaseolus vulgaris*, L.) plants. *Ecotoxicol. Environ. Saf.*, 100 69-75.
- 35 ISTA. (1993) International Rules for Seed Testing. Seed Sci. and Technol. 2 supplement, 25-254.
- 36 ISTA. (1999) International Rules for Seed Testing. Seed Sci. and Technol. 2 supplement, 333 0251-0952.
- 37 Khaleel R. I., Ismail N., and Ibrahim M. H. (2013) The impact of waste water treatments on seed germination and biochemical parameter of Abelmoschus Esculentus L. Procedia Soc. Behav. Sci., 91 453-460.

38 Mabhaudhi T., and Modi A. T. (2011) Can hydro-priming improve germination speed, vigour and emergence of maize landraces under water stress?. J. Agric. Sci. Technol., 1 20-28.

- 39 Maron J. L., Waller L. P., Hahn M. A., Diaconu A., Pal R. W., Muller-Scharer H., Klironomos J. N., and Callaway R. M. (2013) Effects of soil fungi, disturbance and propagule pressure on exotic plant recruitment and establishment at home and abroad. J. Ecol., 101 (4) 924-932.
- 40 Mehboob W., Rehman H., Basra S. M. A., and Afzal I. (2011) Role of seed priming in improving performance of spring maize. In: Proceedings of the international seminar on crop management: issues and options, *Univ. of Agric. Faisalabad, Pakistan* pp, 55.
- 41 Muhammad A. I. (2015) Improving germination and seedling vigour of cowpea (Vigna unguiculata L.) with different priming techniques. *Am.-Eurasian j. agric. environ. sci.*, 15 (2) 265- 270.
- 42 Nikishina T. V., Popov A. S., Kolomeitseva G. L., and Golovkin B. N. (2001) Effect of cryoconservation on seed germination of rare tropical Orchids. *Russ. J. Plant Physiol.*, 48 (6) 810-815.
- 43 Nouman W., Siddiqui M. T., and Basra S. M. A. (2012) *Moringa oleifera* leaf extract: an innovative priming tool for rangeland grasses. *Turk. J. Agric. For.*, 35 65-75.
- 44 Patane C., Saita A., Tubeileh A., Cosentino S. L., and Cavallaro V. (2016) Modeling seed germination of unprimed and primed seeds of sweet sorghum under PEG-induced water stress through the hydrotime analysis. *Acta Physiol. Plant.*, 38 (5) 1-12.
- 45 Phiri C. (2010) Influence of Moringa oleifera leaf extracts on germination and early seedling development of major cereals. Agric. Biol. J. N. Am., 1 (5) 774-777. <u>http://www.scihub.org/ABJNA</u>
- 46 Rady M. M., Gamal F., Mohamed A. M., and Yasmin H. M. (2015) Integrated application of salicylic acid and Moringa oleifera leaf extract alleviates the salt-induced adverse effects in common bean plants. J. Agric. Technol., 11 (7) 1595-1614.
- 47 Rehman H., Nawaz M. Q., Basra S. M. A, Afzal I., Yasmeen A., and Hassan F. U. (2014) Seed priming influence on early crop growth, phenological development and yield performance of linola (*Linum usitatissimum L.*). J. Integr. Agric., 13 (5) 990-996.
- 48 Snedecor G. W., and Cochran W. G. (1992) Statistical Methods. 8th ed., Iowa State Univ., Press, Ames. Iowa, USA.
- 49 Wahid A., and Farooq M. (2012) Is seed invigoration economical and practical?. J. Agric. Soc. Sci., 8 (2) 79-80.
- 50 Wang H. Y., Chen C. L., and Sung J. M. (2002) Both warm water soaking and solid priming treatments enhance anti-oxidation of bitter gourd seeds germinated at sub-optimal temperature. *Seed Sci. Technol.*, 31 47-56.
- 51 Saber A. F., Sayed M., Tolba M. S., Kamal A. M., Hassanien R., and Ahmed M. (2021) A Facile Method for Preparation and Evaluation of the Antimicrobial Efficiency of Various Heterocycles Containing Thieno[2,3-d]Pyrimidine. *Synth. Commun.*, 51 (3) 398-409.
- 52 Ahmed M., Sayed M., Saber A. F., Hassanien R., Kamal El-Dean A. M., and Tolba M. S. (2020) Synthesis, Characterization, and Antimicrobial Activity of New Thienopyrimidine Derivatives. *Polycycl. Aromat. Compd.*, Accepted Manuscript (DOI: 10.1080/10406638.2020.1852587).
- 53 Kamal El-Dean A. M., Zaki R. M., Radwan S. M., and Saber A. F. (2017) Synthesis, Reactions and Spectral Characterization of Novel Thienopyrazole Derivatives. *Eur. Chem. Bull.*, 6 (12) 550–553.
- 54 Zaki R. M., Kamal El-Dean A. M., Radwan S. M., and Saber A. F. (2019) Efficient synthesis, reactions and spectral characterization of novel pyrazolo[4',3':4,5]thieno[3,2-d]pyrimidine derivatives and their related heterocycles. *Heterocycl. Commun.*, 25 (1) 39–46.
- 55 Saber A. F., Zaki R. M., Kamal El-Dean A. M., and Radwan S. M. (2020) Synthesis, reactions and spectral characterization of some new biologically active compounds derived from thieno[2,3-c]pyrazole-5-carboxamide. J. Heterocyclic Chem., 57 (1) 238–247.
- 56 Zaki R. M., El-Dean A. M. K., Radwan S. M., and Saber A. F. (2018) A Convenient Synthesis, Reactions and Biological Activity of Some New 6H-Pyrazolo[4',3':4,5]thieno[3,2-d][1,2,3]triazine Compounds as Antibacterial, Anti-Fungal and Anti-Inflammatory Agents. J. Braz. Chem. Soc., 29 2482-2495.
- 57 Saber A. F., Kamal El-Dean A. M., Redwan S. M., and Zaki R. M. (2020) Synthesis, spectroscopic characterization, and in vitro antimicrobial activity of fused pyrazolo[4',3':4,5]thieno[3, 2-d]pyrimidine. J. Chin. Chem. Soc., 67 (7) 1239-1246.
- 58 Abd-Ella A. A., Metwally S. A., Abdul-Malik M. A., El-Ossaily Y. A., Abd Elrazek F. M., Aref S. A., Naffea Y. A., and Abdel-Raheem Sh. A. A. (2022) A review on recent advances for the synthesis of bioactive pyrazolinone and pyrazolidinedione derivatives. *Curr. Chem. Lett.*, Accepted Manuscript (DOI: 10.5267/j.ccl.2022.2.004).



© 2022 by the authors; licensee Growing Science, Canada. This is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC-BY) license (http://creativecommons.org/licenses/by/4.0/).