

## Synthesis of 3-(2-(substituted-(trifluoromethyl)phenylamino)acetyl)-2H-chromen-2-one derivatives as new anticancer agents

Anees Pangal<sup>a</sup>, Yusufi Mujahid<sup>a</sup>, Bajarang Desai<sup>a</sup>, Javed A. Shaikh<sup>a</sup> and Khursheed Ahmed<sup>a\*</sup>

<sup>a</sup>Advanced Scientific Research Laboratory (ASR LAB.), Department of Chemistry, Abeda Inamdar Senior College of Arts, Science & Commerce, Camp, Pune – 411001, India

### CHRONICLE

#### Article history:

Received June 2, 2021  
Received in revised form  
June 18, 2021  
Accepted August 31, 2021  
Available online  
August 31, 2021

#### Keywords:

Coumarin  
Anticancer  
MCF-7  
HeLa  
SCC-40  
SRB Assay

### ABSTRACT

Under solvent free conditions and in presence of a base 3-(2-(substituted-(trifluoromethyl)phenylamino)acetyl)-2H-chromen-2-one derivatives were synthesized by grinding technique. Structural investigations were carried out with IR studies, HRMS, <sup>1</sup>HNMR and <sup>13</sup>CNMR. The compounds were checked for their *in vitro* anticancer activities against three different human cancer cell lines viz human breast cancer cell line (MCF-7), human cervical cancer cell line (HeLa) and human oral squamous cell carcinoma (SCC-40) using SRB method. All the title compounds showed low toxicity towards non-malignant PBMC cells indicating their tumour selectivity. The compounds exhibited good *in vitro* anti-proliferative potency at lower concentrations against HeLa and MCF-7 cell lines and remain moderately active against SCC-40.

© 2022 by the authors; licensee Growing Science, Canada.

## 1. Introduction

Anti-cancer drugs are meant to target abnormally dividing cell through inhibition of cell division.<sup>1</sup> antiproliferative activity of anti-cancer compounds involve diverse mechanism and accordingly such compounds are called as DNA intercalating agents (*e.g.* adriamycin), DNA cross-linking agents (*e.g.* cis-platin), topoisomerase inhibitors (*e.g.* camptothecins), cytoskeleton-disrupting agents (*e.g.* vinblastin), tyrosine kinase inhibitors (imatinib) and antimetabolites (*e.g.* mercaptopurine). One of the biggest challenges in chemotherapy is the collateral damage to the normal cells by chemotherapeutic agents and consequent severe side effects and selective activity against cancer cell is desirable attribute in all anti-cancer compounds.<sup>2</sup>

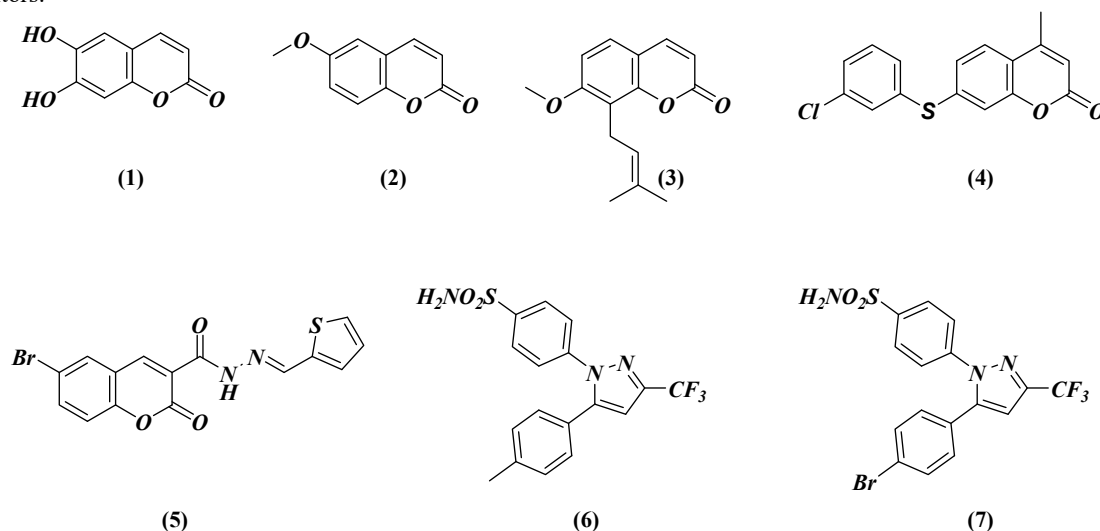
Coumarins are heterocyclic compounds from family of lactones with 1-benzopyran-2-one system and they can be isolated from essential oils, green tea, fruits and they can be synthesized in the laboratory.<sup>3,4</sup> Coumarin is a versatile phytochemical and its derivatives are known to exhibit antibacterial and antimicrobial activities with low toxicity.<sup>5,6,7</sup> Coumarin derivatives have also been reported for anticoagulant and anti-inflammatory,<sup>8</sup> anti-HIV,<sup>9</sup> antioxidant,<sup>10</sup> antiallergic,<sup>11</sup> anticancer,<sup>12</sup> antiproliferative<sup>13</sup> and antiviral<sup>14</sup> activities and these pharmacological properties depend substituent.<sup>15</sup> Coumarins could exert their anticancer activity by different mechanisms either by inhibiting the telomerase enzyme<sup>16</sup> and down regulating oncogene expression<sup>17</sup> or by inducing the caspase-9 mediated apoptosis. Additionally, researchers showed that coumarins are able to suppress cancer cell proliferation by arresting cell cycle in G<sub>0</sub>/G<sub>1</sub>,<sup>18</sup> G<sub>2</sub>/M

\* Corresponding author.

E-mail address: [khursheedahmed@azamcampus.org](mailto:khursheedahmed@azamcampus.org) (K. Ahmed)

© 2022 by the authors; licensee Growing Science, Canada  
doi: 10.5267/j.ccl.2021.008.004

phases<sup>19</sup> and through affecting the p- glycoproteins of the cancer cells.<sup>20</sup> It was also reported that hydroxycoumarins might exert their anticancer activity by generating free radical species causing oxidative stress leading to pro-apoptotic effects.<sup>21</sup> It was proven that the - $\gamma$ -lactone ring of the coumarinic system has a fundamental role in both the generation and stabilization of such species as well as in the pro-apoptotic action of hydroxycoumarins.<sup>22</sup> Moreover, the antiproliferative activity of 7-hydroxycoumarin derivatives could be due to their effect on the mitochondrial thiols.<sup>23</sup> Many natural and synthetic coumarin compounds (**Scheme 1**) have shown promising potentials to be anticancer agents. Esculetin (6,7-dihydroxycoumarin, **1**) and Scopoletin (6-methoxy-7-hydroxycoumarin, **2**) which are, typical naturally occurring coumarins displayed antiproliferative effects in human leukaemic cells.<sup>24</sup> Compound **1** is known to inhibit proliferation and induce cell death in many forms of human cancer cells and is considered as a promising chemotherapeutic agent. Recently, Cho *et al.* reported antiproliferative effect of **2** on the growth of oral squamous cell carcinoma cell lines.<sup>25</sup> Zhang *et al.* reported potent cytotoxicity of compound **3** against human hepatocellular carcinoma cells.<sup>26</sup> Chen *et al.* reported compound **4** with remarkable anti-proliferative activity against human prostate cancer and human lung cancer cell line.<sup>27</sup> Coumarin derivative **5** synthesized by Nasr *et al.*<sup>28</sup> was found to possess powerful growth inhibitory activity against human hepatocellular carcinoma and human pancreatic carcinoma cell lines. In addition coumarin-containing molecules have been shown to reverse the multidrug resistance (MDR) in various<sup>29</sup> which underlines the future prospect of coumarin compounds against resistant cancer. The fluorine-containing organic compounds are particularly appreciated in pharmaceutical, agricultural and materials sciences. In recent years, the trifluoromethyl group has attracted more attention, and many trifluoromethylated compounds have been found to possess special activities.<sup>30</sup> Due to its stereoelectronic property and lipophilicity,<sup>31</sup> the introduction of trifluoromethyl groups into bioactive molecules has led to the synthesis of new molecules with remarkable therapeutic potential,<sup>32</sup> for example Celecoxib **6** and SC-558 **7** exhibited potent chemo preventive activity as COX-2 inhibitors.



**Scheme 1.** Reported Coumarin Derivatives

The sulforhodamine B (SRB) assay is employed for the determination of cell density which is based on the measurement of cellular protein content and the method relies on the stoichiometric binding of SRB with proteins under slightly acidic conditions.<sup>33-34</sup> Further, molecular docking is a well-known tool to explore binding interaction between drug and receptor. The interaction of a synthesized drug molecules with Cyclooxygenase (COX) target via docking and their relative stabilities have been evaluated using the binding affinities.<sup>35</sup> Cyclooxygenase (COX) enzymes, also known as prostaglandin-endoperoxide synthase (PTGS), catalyze the metabolic conversion of arachidonic acid (AA) to prostaglandins (PGs) that play an important role in inflammation.<sup>36</sup> COX-2 is a membrane-bound, short-living, rate-limiting enzyme is a known target for the treatment of inflammation. COX-2 expression is negligible in normal cells but it is expressed frequently at the tumorigenic nests in many types of cancers including adenocarcinoma, squamous cell carcinoma (SCC) and hepatocellular carcinoma.<sup>37</sup> Recently, benzopyran derivatives were investigated as potent COX-2 inhibitors and more specifically, coumarin derivatives, as a class of benzopyrans were proved to possess potent anti-inflammatory effects and thus were evaluated as COX-2 inhibitors.<sup>38</sup> Our interest in the identification of new COX-2 inhibitors prompted us to explore the use of coumarin framework for the design of this type of inhibitors.<sup>38</sup> Researchers have condensed 3-(2-bromo acetyl) coumarin with aniline in ethanol at reflux temperature for 15–30 min which resulted in the formation of 3-(2-(phenylanilino)acetyl)-2H-chromen-2-one but the yield of the products is around 70–75%.<sup>39</sup> Though, these methodologies are quite simple and useful but they need prolonged reaction time and give moderate yields of the products. Hence, there is a requirement of alternative procedures with mild reaction conditions and better yields. In the present work we report the synthesis of 3-(2-(phenylamino)acetyl)-2H-chromen-2-ones under solvent-free conditions. The synthesized compounds were tested for their anticancer potential against three cancer cell lines such as human breast cancer cell line (MCF-7), human cervical cancer cell line (HeLa) and human oral squamous cell carcinoma (SCC-40) by SRB method.

## 2. Results and Discussion

### 2.1 Spectral Characterization

The HRMS (EI) spectra of **3ACOT**, **3ACMT**, **3ACPT** and **3ACDT** showed major peaks corresponding to expected M+1 fragment at 348.0837, 348.0838, 348.0842 and 438.0535 respectively. The IR spectra of these analogs showed characteristic peaks for lactone carbonyl, -NH and >C=O (amide). The IR spectrum of all the analogs showed one peak in between 3020 to 3047 cm<sup>-1</sup> for -NH. The peak at 1714 to 1724 cm<sup>-1</sup> is due to >C=O (lactone moiety) and that at 1675 to 1686 cm<sup>-1</sup> is due to >C=O (ketonic moiety). The <sup>1</sup>H-NMR spectrum of the synthesized analogs showed -CH<sub>2</sub> protons at 4.7 ppm. The olefin proton on C4 of coumarin ring appeared as a sharp singlet at 8.6 ppm. The aromatic hydrogen atoms were located in the range of 6.7 to 7.7 ppm. On the other hand, the protons of -NH groups appeared as singlet at 2.17 ppm. The <sup>13</sup>C-NMR of all these analogs exhibited signals from aromatic carbon atoms at 109.3 to 149.6 ppm and characteristic peaks of ketonic carbonyl and lactone carbons at about 185.8 to 192.7 and 155.4 to 158.9 ppm respectively. In all these compounds the aliphatic carbon i.e. -CH<sub>2</sub> signals were found in the range of 35 to 54 ppm.

- i) **3-(2-(2-(trifluoromethyl)phenylamino)acetyl)-2H-chromen-2-one (3ACOT)**: Pale yellow powder, yield 90 %, MP 205-206 °C, IR(cm<sup>-1</sup>): 3020 (NH), 1724 (C=O of lactone), 1685 (C=O of ketone); <sup>1</sup>HNMR (500 MHz, CDCl<sub>3</sub>): δ 2.170 (s, 1H), 4.751 (s, 2H), 7.366 to 7.722 (m, 8H), 8.635 (s, 1H, =CH); <sup>13</sup>C NMR (500MHz, CDCl<sub>3</sub>): 35, 116.9, 118.1, 122, 122.1, 122.1, 122.1, 125.3, 125.3, 125.3, 125.3, 130.4, 135, 149.1, 155.4, 185.8, 188.8; HRMS (EI): C<sub>18</sub>H<sub>12</sub>F<sub>3</sub>NO<sub>3</sub>: 348.0837
- ii) **3-(2-(3-(trifluoromethyl)phenylamino)acetyl)-2H-chromen-2-one (3ACMT)**: Yellow powder, yield 92 %, MP 198-200 °C, IR(cm<sup>-1</sup>): 3047 (NH), 1714 (C=O of lactone), 1675 (C=O of ketone); <sup>1</sup>HNMR (500 MHz, CDCl<sub>3</sub>): δ 2.166 (s, 1H, -NH), 4.746 (s, 2H, -CH<sub>2</sub>), 6.803 (m, 1H), 6.984 (dd, 1H), 6.898 (dd, 1H), 6.85 (dd, 1H), 7.047 to 7.283 (m, 4H), 8.638 (s, 1H, =CH); <sup>13</sup>CNMR (500MHz, CDCl<sub>3</sub>): 54, 109.3, 114.8, 116.8, 121.9, 123.3, 124.6, 125.2, 125.3, 129.7, 130.7, 130.7, 131.6, 135.1, 147.7, 149.6, 158.9, 192.7; HRMS (EI): C<sub>18</sub>H<sub>12</sub>F<sub>3</sub>NO<sub>3</sub>: 348.0838
- iii) **3-(2-(4-(trifluoromethyl)phenylamino)acetyl)-2H-chromen-2-one (3ACPT)**: Brown powder, yield 92%, MP 210-212 °C, IR (cm<sup>-1</sup>): 3037 (NH), 1723 (C=O of lactone), 1686 (C=O of ketone); <sup>1</sup>HNMR (500 MHz, CDCl<sub>3</sub>): δ 2.166 (s, 1H, -NH), 4.747 (s, 2H, -CH<sub>2</sub>), 6.708 (q, 2H), 7.720 (q, 2H), 7.263 to 7.440 (m, 4H), 8.628(s, 1H, =CH); <sup>13</sup>CNMR (500MHz, CDCl<sub>3</sub>): 53.8, 112.2, 119.3, 121.8, 122.1, 124.1, 125.3, 126.7, 127.2, 127.2, 130.4, 133, 135.3, 149.6, 158.9, 192.6; HRMS (EI): C<sub>18</sub>H<sub>12</sub>F<sub>3</sub>NO<sub>3</sub>: 348.0842
- iv) **3-(2-(3,5-bis(trifluoromethyl)phenylamino)acetyl)-2H-chromen-2-one (3ACDT)**: Yellow powder, yield 90%, MP 204-206 °C; IR(cm<sup>-1</sup>): 3024 (NH), 1723 (C=O of lactone), 1684 (C=O of ketone); <sup>1</sup>HNMR (500 MHz, CDCl<sub>3</sub>): δ 2.171 (s, 1H, -NH), 4.750 (s, 2H, -CH<sub>2</sub>), 7.063 (s, 2H), 7.195 (d, 1H), 7.367 to 7.723 (m, 4H), 8.636 (s, 1H, =CH); <sup>13</sup>CNMR (500MHz, CDCl<sub>3</sub>): 53.7, 112.3, 116.9, 116.9, 118.1, 122.1, 125.3, 125.4, 130.4, 130.4, 130.7, 130.7, 135.1, 147.3, 149.6, 158.9, 191.9; HRMS (EI): C<sub>19</sub>H<sub>11</sub>F<sub>6</sub>NO<sub>3</sub>Na: 438.0535

### 2.2 Anti-cancer Activity

The anti-proliferative activities of four synthesized coumarin derivatives were tested against human breast cancer cell line (MCF-7), human cervical cancer cell line (HeLa) and human oral squamous cell carcinoma (SCC-40) and the results are depicted in **Table 1**. The results for are each compound are expressed as the percent growth (GP %) at different concentrations of drug compounds and expressed in µg/ml. Adriamycin (ADR) is used as standard. The growth curves of different cancer cell lines and normal human peripheral blood mononuclear cells (PBMC) are shown in **Fig. 1** to **Fig. 4**. The activities of test compounds were compared with those of non-malignant normal human peripheral blood mononuclear cells (PBMCs). The results reveal that all the compounds show weak cytotoxicities against normal cells PBMC and thus the findings comply that all compounds were selective against cancer cells.

**Table 1.** Average values of percentage control growth of 3 cell lines and normal human peripheral blood mononuclear cells (PBMC) at different concentrations in µg/ml

Cell Lines Drug Concentrations (µg/ml)	MCF-7				HeLa				SCC-40				PBMC			
	10	20	40	80	10	20	40	80	10	20	40	80	10	20	40	80
<b>3ACOT</b>	101.3	97.2	104.1	76.6	47.2	45.4	32.3	16.0	85.4	87.8	63.0	24.2	99.67	93.43	90.53	88.17
<b>3ACMT</b>	97.7	103.4	97.9	76.4	48.8	49.1	26.8	-12.5	98.1	95.8	78.9	27.6	97.22	93.95	87.62	80.17
<b>3ACPT</b>	95.5	89.5	85.0	86.2	73.9	73.8	42.2	20.9	93.3	83.4	66.0	37.7	95.29	89.27	83.72	79.98
<b>3ACDT</b>	92.1	78.3	48.8	11.1	101.5	101.4	60.8	47.1	86.4	72.1	-1.6	-63.1	97.44	90.25	85.04	79.78
<b>ADR</b>	-48.5	-48.1	-40.6	-14.2	-56.9	-58.1	-62.9	-49.4	-84.6	-84.8	-86.7	-77.9	-	-	-	-

From the observed data it is noted that all the synthesized compounds show appreciable activity against human cervical cancer cell line (HeLa) compared to other two cell lines. From the anti-proliferative screening data, it is evident that at higher concentrations, the compounds show moderate activities.

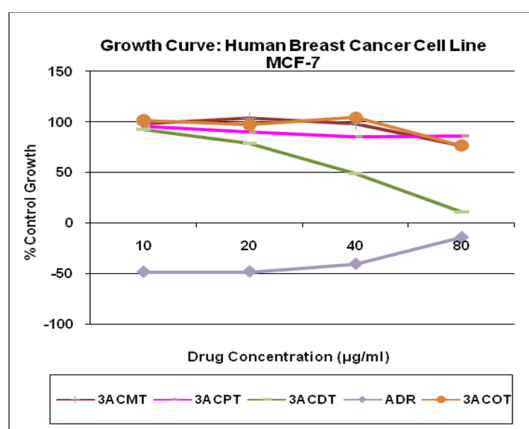


Fig. 1. Growth Curve: Human Breast Cancer Cell Line (MCF-7)

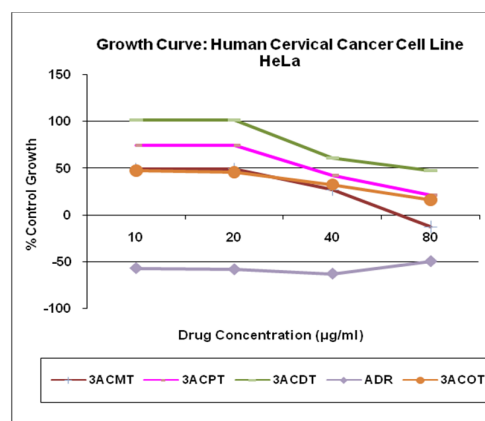


Fig. 2. Growth Curve: Human Cervical Cancer Cell Line (HeLa)

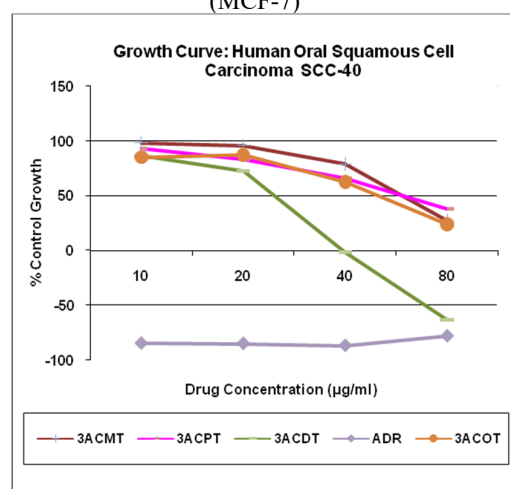


Fig. 3. Growth Curve: Human Oral Squamous Cancer Cell Line (SSC-40)

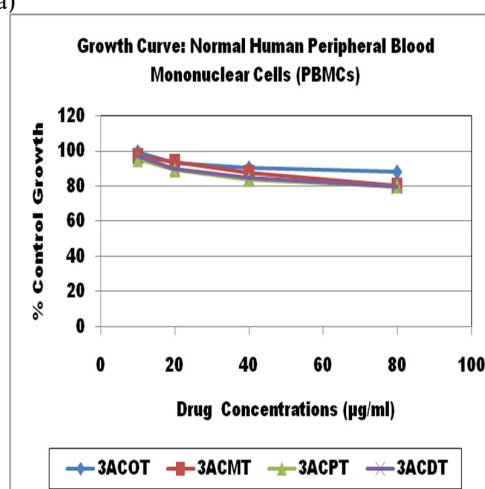


Fig. 4. Growth Curve: Normal Human Peripheral Blood Mononuclear Cells (PBMCs)

Out of the four synthesized derivatives, 3ACDT was found to be most effective against MCF-7 cell lines at 10 µg/ml concentration. 3ACOT and 3ACMT expresses best activity against HeLa cell line at 10 µg/ml concentration. 3ACOH and 3ACMT shows the average cell growth percents of 47.2 and 48.8 respectively against HeLa cancer lines. However, amongst the synthesized coumarin derivatives 3ACOT and 3ACDT were the most active with the average cell growth percents of 85.4 and 86.4 respectively against SCC-40 cell lines. Three compounds show significant anti-proliferative activity against human cervical cancer cell line (HeLa) at lower concentrations in between 47.2 to 73.9 except 3ACDT showing percent growth of 101.5. These compounds demonstrated cytotoxic effect on MCF-7 and SCC-40 cancer cell lines with the average cell growth percent values of 92.1 to 101.3 and 85.4 to 93.3 respectively at the same concentrations. Thus, the synthesized compounds are exceedingly active against HeLa and SCC-40 cell lines and moderately active against MCF-7.

The parameter GI50 was calculated using the graph of % control inhibition values and drug concentrations. The results are shown in Table 2.

Table 2. GI50 values (µg/ml) for 3 cell lines

Cell Lines	MCF-7	HeLa	SSC-40
Codes	GI50*	GI50*	GI50*
3ACOT	53.6	<10	>80
3ACMT	61.5	13.8	>80
3ACPT	63.1	40.8	>80
3ACDT	25.4	70.6	44.1
ADR	<10	<10	<10

GI50 values are calculated for the 50% growth inhibition of cells. For getting the idea of the activity of the compounds the GI50 value of  $\leq 10 \mu\text{g/ml}$  is considered as a good activity. It is observed that the GI50 value for 3ACMT is  $13.8 \mu\text{g/ml}$  against HeLa and other derivatives exhibit the GI50 values  $40.8$  and  $70.6 \mu\text{g/ml}$  for 3ACPT and 3ACDT respectively. The derivative 3ACOT shows the GI50 value comparable to standard ADR which is less than  $10 \mu\text{g/ml}$ . In terms of GI50 values, all the derivatives are active against human cervical cancer cell line (HeLa). 3ACDT is found to be most active against MCF-7 with GI50 value of  $25.4 \mu\text{g/ml}$ .

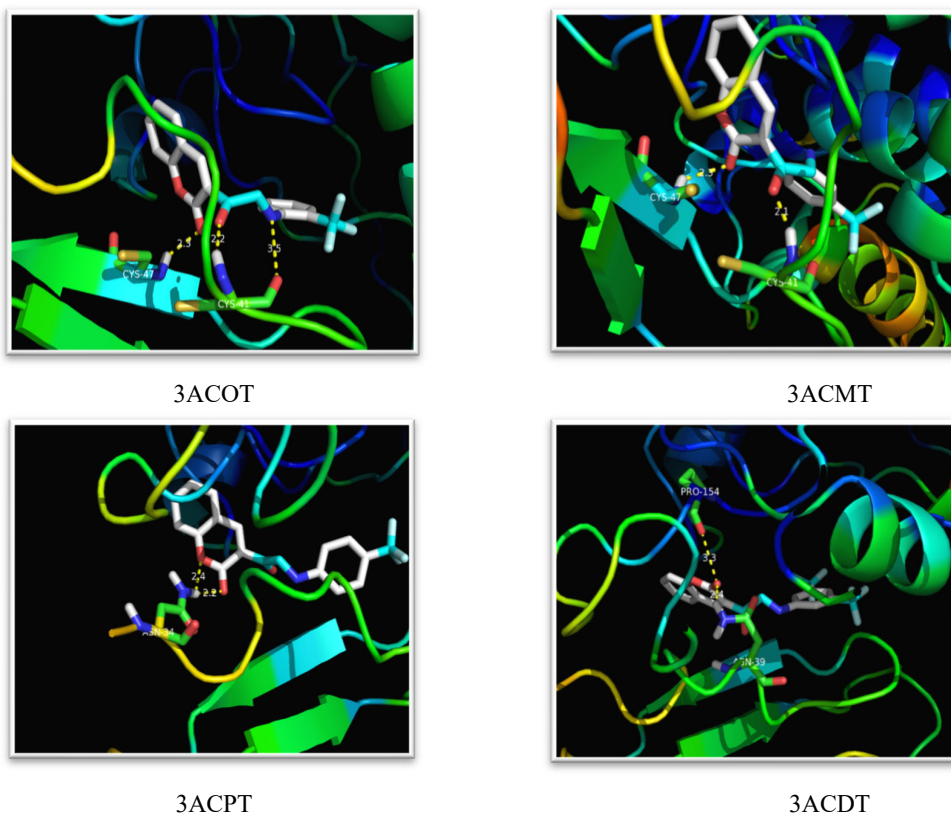
### 2.3 Molecular Docking

Molecular docking study was performed to check the binding affinities of synthesized analogues with the Cyclooxygenase active site residues of COX-2 enzyme (PDB ID: 6COX) using the Auto Dock 4.2.3 software. A grid box covering the cyclooxygenase active site residues of the target protein was generated to induce the most effective conformational state of docking. Docking grid box size was set to  $46 \times 46 \times 46 \text{ \AA}$  dimension and focused at  $30.77, -1.64, 25$  of X, Y and Z coordinates. Present docking study affirmed that all the four derivatives fit favorably into the cyclooxygenase active site of COX-2 displaying H-bonding interactions with CYS-41, CYS-47, ASN-34 and PRO-154 amino acid residues. The estimated free energy values of binding and the interacting amino acid residues are given in **Table 3**.

**Table 3.** Binding energy and the interacting amino acid residues

Sr. No.	Compounds	B.E. kcal/mole	Binding amino acid Residue	Bond Lengths $\text{\AA}$
1	3ACOT	-9.6	CYS-41, CYS-47	2.2, 3.5, 2.3
2	3ACMT	-9.6	CYS-41, CYS-47	2.1, 2.3
3	3ACPT	-9.3	ASN-34	2.2, 2.4
4	3ACDT	-10	ASN-34, PRO-154	2.4, 3.3

The docking ribbon structures of 6COX protein with respective compounds are given in **Fig. 5**. The best binding energy was exhibited by 3ACDT followed by 3ACOT, 3ACTMT and 3ACPT respectively. 3ACDT shows hydrogen bonding interactions with two amino acid residues, viz. ASN-34 and PRO-154 in the protein cavity. The hydrogen bonding distances are  $2.4$  and  $3.3 \text{ \AA}$ . The derivative 3ACOT shows affinity with two amino acid residues CYS-41 and CYS-47 with formation of 3 hydrogen bonds. These interactions cause stabilization of the compounds within the protein cavity. Based on these observations, compound 3ACDT has better stability within the 6COX protein cavity than other derivatives and contemplated to show enhanced anticancer activity.



**Fig. 5.** Binding of four 3-(2-(substituted-(trifluoromethyl)phenylamino)acetyl)-2H-chromen-2-one derivatives into the active site of 6COX assessed by molecular docking



## 2H-chromen-2-one derivatives

**4.3 SRB Assay:** The *in vitro* anticancer testing was carried out using the SRB assay protocols.<sup>41</sup> The test compounds were inoculated at 4 dose levels at 10, 20, 40, 80 µg/ml. Cell lines were counted, cultured and inoculated in 96 well plates. Each experiment was repeated three times. After incubation with different concentrations of test compounds, the cell cultures were stained with SRB dye. After washing with 1% acetic acid the protein bounded dye was extracted using Tris-HCl buffer base (100 µl, 0.01 M, pH 10.4). The optical density was determined on 96-well plate ELISA reader at 540 nm. Cell viability was expressed as a percentage growth of sample compounds with that of the control.

**4.4 Cytotoxicity assay against non-cancerous cells:** The cytotoxicity of synthesized analogues was performed on non-cancerous cells i.e. normal human peripheral blood mononuclear cells (PBMCs). The PBM cells were isolated from whole blood of human healthy volunteers using the procedure available in literature.<sup>42</sup> PBMCs were seeded on 96-well microplates at a density of  $1 \times 10^4$ . Next day the culture medium was removed and cells were exposed to serial dilutions of the synthesized compounds in fresh culture medium. Cell proliferation was studied for 24-48 hours by means of MTT assay in which the yellow tetrazolium salt (MTT) is metabolized by viable cells to purple formazan crystals. The plates were incubated for 3 hours with MTT solution (5 mg/ml). Formazan crystals were dissolved in DMSO<sub>4</sub> and the purple colour developed was observed spectrophotometrically at 570 nm wavelength using Readwell Touch Automatic Elisa Plate Reader (Robonik India Private Limited).

**4.5 In silico Molecular Docking:** Molecular docking studies were carried out on cyclooxygenase active site residues of COX-2 enzyme (PDB ID: 6COX) using the software, Auto Dock 4.2.614. The PDB file (6COX) of human cyclooxygenase enzyme was downloaded from Royal Society Protein Data Bank. The PDB was processed in Discovery Studio for removal of DNA and water molecules to make the binding sites free for interaction with the synthesized compounds. The cleaned up PDB files were used for generation of PDBQT file by adding Kollmann charges and further used for docking studies. The images were created with the help of Pymol Molecular Viewer software.<sup>43</sup>

## References

- Pangal A., Shaikh J. A., and Khan E. M. (2017) Current developments of C3-substituted coumarin hybrids as anti-cancer agents. *Int. J. Pharm. Sci. Rev. Res.*, 42 (1), 161-168.
- Jain P. K., and Joshi H. (2012) Coumarin: Chemical and pharmacological profile. *J. Appl. Pharm. Sci.*, 02 (06), 236-240.
- Batra N., Batra S., Pareek A., and Nagori B. P. (2012) Diverse pharmacological activities of 3-substituted coumarins: A review. *Int. Res. J. of Pharma.*, 3 (7), 24-29.
- Pangal A., Gazge M., Mane V., and Shaikh J. A. (2013) Various pharmacological aspects of coumarin derivatives: A review. *IJPRBS*, 2 (6), 168-194.
- Shridhar R., Sastry, Reddy C. V., Vaidya N. K., Moorthy S. R., Reddy G. S., and Thapar G. S. (1978) Antimicrobial agents: Synthesis and antibacterial activity of some new 4-[2-(heteroaryl) vinyl]coumarins. *Ind. J. Chem.*, 16B, 704-708.
- Gadaginmath, Guru S., Joshi, and Raghvendra G. (1995) Synthesis and antimicrobial activity of benimidazolyl / coumarinylmethoxyindoles. *Ind. J. Chem.*, 34B, 120-124.
- Olayinka O. A., and Nwinyi O. C. (2010) Microwave-assisted synthesis and evaluation of antimicrobial activity of 3-{3-(s-aryl and s-heteroaromatic)acryloyl}-2H-chromen-2-one derivatives. *J. Heterocyclic Chem.*, 47, 179-187.
- Rajeshwar R. V., Srimanth K., and Kumar V. P. (2004) Synthesis of 3-[3-substituted thio]-7H-1,2,4-triazolo[3,4-b][1,3,4]thiadiazin-6-yl]-2H-1-benzopyran-2-ones and 3-[3-aminoaryl/heterocyclyl]-7H-1,2,4-triazolo[3,4-b][1,3,4]thiadiazin-6-yl]-2H-1-benzopyran-2-ones. *Ind. J. Heter. Chem.*, 14, 141-144.
- Srivastav V. K., Tiwari M., Zhang X., and Yao X-J (2018) Synthesis and antiretroviral activity of 6-acetyl-coumarin derivatives against HIV-1 infection. *Ind. J. Pharm. Sci.*, 80 (1), 108-117.
- Bubols G. B., Vianna D. R., Medina-Reimon A., von Poser G., Lamuela-Raventos R. M., Eifler-Lima V. L., and Garcia S. C. (2013) The antioxidant activity of coumarins and flavonoids. *Mini Rev. Med. Chem.*, 13 (3), 318-334.
- Lin C. L., Hsiao G., Wang C. C., and Lee Y. L. (2016) Imperatorin exerts antiallergic effects in Th2-mediated allergic asthma via induction of IL-10-producing regulatory T cells by modulating the function of dendritic cells. *Pharmacol. Res.*, 110, 111-121.
- Klenkar J., and Molnar M. (2015) Natural and synthetic coumarins as potential anticancer agents. *J. Chem. & Pharma. Res.*, 7 (7), 1223-1238.
- Zhang Y., Zhang Q., Song J., Zhang L., Jiang C., and Zhang H. (2018) Design, Synthesis, and Antiproliferative Evaluation of Novel Coumarin/2-Cyanoacryloyl Hybrids as Apoptosis Inducing Agents by Activation of Caspase-Dependent Pathway. *Molecules*, 23, 1972-1991.
- Hassan M. Z., Osman H., Ali M. A., and Ahsan M. J. (2016) Therapeutic potential of coumarins as antiviral agents. *Eur. J. Med. Chem.*, 123, 236-255.
- Kostova I (2006) Synthetic and natural coumarins as antioxidants. *Mini Rev. Med. Chem.*, 6 (4), 365-374.
- Wu X. Q., Huang C., Jia Y. M., Song B. A., Li J., and Liu X. H. (2014) Novel coumarin-dihydropyrazole thio-ethanone derivatives: design, synthesis and anticancer activity. *Eur. J. Med. Chem.*, 74, 717-725.
- Vianna D. R., Hamerski L., Figueiró F., Bernardi A., Visentin L. C., Pires E. N., Teixeira H. F., Salbego C. G., Eifler-Lima V. L., Battastini A. M., von Poser G. L., and Pinto A. C. (2012) Selective cytotoxicity and apoptosis induction in glioma cell lines by 5-oxygenated-6,7-methylenedioxy coumarins from *Pterocaulon* species. *Eur. J. Med. Chem.*, 57, 268-274.
- Chen Y., Liu H. R., Liu H. S., Cheng M., Xia P., Qian K., Wu P. C., Lai C. Y., Xia Y., Yang Z. Y., Morris-Natschke S. L., and Lee K. H. (2012) Antitumor agents 292. Design, synthesis and pharmacological study of S- and O-substituted 7-mercapto- or hydroxy-coumarins and chromones as potent cytotoxic agents. *Eur. J. Med. Chem.*, 49, 74-85.



19. Fong W. F., Shen X. L., Globisch C., Wiese M., Chen G. Y., Zhu G. Y., Yu Z. L., Tse A. K., and Hu Y. J. (2008) Methoxylation of 3',4'-aromatic side chains improves P-glycoprotein inhibitory and multidrug resistance reversal activities of 7,8-pyrano coumarin against cancer cells. *Bioorg. Med. Chem.*, 16, 3694-3703.
20. Zhou T., Shi Q, Bastow K. F., and Lee K. H. (2010) Antitumor agents 286. Design, synthesis, and structure-activity relationships of 3'R,4'R-disubstituted-2',2'-dimethyldihydropyrano[2,3-f]chromone (DSP) analogues as potent chemosensitizers to overcome multidrug resistance. *J. Med. Chem.*, 53, 8700-8708.
21. Vázquez R., Riveiro M. E., Vermeulen M., Alonso E., Mondillo C, Facorro G., Piehl L., Gómez N., Moglioni A., Fernández N., Baldi A., Shayo C., and Davio C. (2012) Structure-anti-leukemic activity relationship study of ortho-dihydroxycoumarins in U-937 cells: key role of the  $\delta$ -lactone ring in determining differentiation-inducing potency and selective pro-apoptotic action. *Bioorg. Med. Chem.*, 20 (18), 5537-5549.
22. Riveiro M. E., De Kimpe N., Moglioni A., Vázquez R., Monczor F., Shayo C., and Davio C. (2010) Coumarins: old compounds with novel promising therapeutic perspectives. *Curr. Med. Chem.*, 17 (13), 1325-1338.
23. Molaverdi F., Khoobi M., Emami S., Alipour M., Firuzi O., Foroumadi A., Dehghan G., Miri R., Shaki F., Jafarpour F., and Shafiee A. (2013) Polyoxygenated cinnamoylcoumarins as conformationally constrained analogs of cytotoxic diarylpentanoids: synthesis and biological activity. *Eur. J. Med. Chem.*, 68, 103-110.
24. Dai H., Huang M., Qian J., Liu J., Meng C., Li Y., Ming G., Zhang T., Wang S., Shi Y., Yao Y., Shushan G., Zhang Y., and Ling Y. (2019) Excellent antitumor and antimetastatic activities based on novel coumarin/pyrazole oxime hybrids. *Eur. J. Med. Chem.*, 166, 470-479.
25. Cho J. H., Shin J., Cho J., Choi Y. H., Shim J. H., and Chae J. (2015) Esculetin (6,7-dihydroxycoumarin): a potential cancer chemopreventive agent through suppression of Sp1 in oral squamous cancer cells. *Int. J. Oncol.*, 46, 265-271.
26. Zhang L., Jiang G., Yao F., He Y., Liang G., Zhang Y., Hu B., Wu Y., Li Y. and Liu H. (2012) Growth inhibition and apoptosis induced by osthole, a natural coumarin in hepatocellular carcinoma. *PLoS One.*, 7 (5), e37865.
27. Chen Y., Liu H. R., Liu H. S., Cheng M., Xia P., Qian K., Wu P. C., Lai C. Y., Xia Y., Yang Z. Y., Morris-Natschke S. L., and Lee K. H. (2012) Antitumor agents 292. Design, synthesis and pharmacological study of S- and O-substituted 7-mercapto- or hydroxycoumarins and chromones as potent cytotoxic agents. *Eur. J. Med. Chem.*, 49, 74-85.
28. Nasr T., Bondock S., and Youns M. (2014) Anticancer activity of new coumarin substituted hydrazide-hydrazone derivatives. *Eur. J. Med. Chem.*, 76, 539-548.
29. Bisi A., Cappadone C., Rampa A., Farruggia G., Sargenti A., Belluti F., Di Martino R.M.C., Malucelli E., Meluzzi A., Iotti S., and Gobbi S. (2017) Coumarin derivatives as potential antitumor agents: Growth inhibition, apoptosis induction and multidrug resistance reverting activity. *Eur. J. Med. Chem.*, 127, 577-585.
30. Zhang C. (2014) Recent advances in trifluoromethylation of organic compounds using Umemoto's reagents. *Org. Biomol. Chem.*, 12, 6580-6589.
31. Betageri R., Zhang Y., Zindel R. M., Kuzmich D., Kirrane T. M., Bentzien J., Cardozo M., Capolino A. J., Fadra T. N., Nelson R. M., Paw Z., Shih D. T., Shih C. K., Zuvela-Jelaska L., Nabozny G., and Thomson D.S. (2005) Trifluoromethyl group as a pharmacophore: effect of replacing a -CF<sub>3</sub> group on binding and agonist activity of a glucocorticoid receptor ligand. *Bioorg. Med. Chem. Lett.*, 15 (21), 4761-4769.
32. Lu X. Y., Wang Z. C., Ren S. Z., Shen F. Q., Man R. J., and Zhu H. L. (2016) Coumarin sulfonamides derivatives as potent and selective COX-2 inhibitors with efficacy in suppressing cancer proliferation and metastasis. *Bioorg. Med. Chem. Lett.*, 26 (15), 3491-3498.
33. Vichai V. and Kirtikara K. (2006) Sulforhodamine B colorimetric assay for cytotoxicity screening. *Nat. Protoc.*, 1 (3), 1112-1116.
34. Esteban A., Orellana, and Kasinski A. L. (2016) Sulforhodamine B (SRB) Assay in cell culture to investigate cell proliferation. *Bio. Protoc.*, 6 (21), 1984-1993.
35. Rajeswar R. V., and Kumar R. V. (2002) Synthesis of 3-(1-aryl-2-mercaptoimidazolyl)-2H-1-benzopyran-2-one. *Ind. J. Chem.*, 41B, 415-418.
36. George S., Kumaran S., Chandran M., Gangwar P., and Gururagavan M. (2012) Docking studies of novel coumarin derivatives as arylamine N-acetyltransferase 2 inhibitors. *Asian J. Pharm. Clin. Res.*, 5 (1), 94-96.
37. Smith, W. L., Urade Y., and Jakobsson P. J. (2011) Enzymes of the cyclooxygenase pathways of prostanoid biosynthesis. *Chem. Rev.*, 111, 5821-5865.
38. Hashemi G. N., Najafi M., Salehi E., Farhood B., and Mortezaee K. (2019) Cyclooxygenase-2 in cancer: A review. *J. Cell Physiol.*, 234 (5), 5683-5699.
39. Dawood D. H., Batran R. Z., Farghaly T. A., Khedr M. A., and Abdulla M. M. (2015) New Coumarin Derivatives as Potent Selective COX-2 Inhibitors: Synthesis, anti-inflammatory, QSAR, and molecular modeling studies. *Arch. Pharm. (Weinheim)*, 348 (12), 875-888.
40. Guravaiah N., and Rao R. V. (2011) Efficient, Stereoselective approach to the synthesis of 3-(1-Phenyl-2-(Z-styrylsulfonyl)-1H-imidazol-4-yl)-2H-chromen-2-ones. *Synth. Commun.*, 41 (8), 1167-1174.
41. Skehan P., Storeng R., Scudiero D., Monks A., McMahon J., Vistica D., Warren J. T., Bokesch H., Kenney S., and Boyd M.R. (1990) New colorimetric cytotoxicity assay for anticancer-drug screening. *J. Natl. Cancer Inst.*, 82 (13), 1107-1112.
42. Jenny M., Klieber M., Zaknun D., Schroecksnadel S., Kurz K., Ledochowski M., Schennach H., and Fuchs D. (2011) *In vitro* testing for anti-inflammatory properties of compounds employing peripheral blood mononuclear cells freshly isolated from healthy donors. *Inflamm Res.*, 60 (2), 127-135.
43. Pangal A., Gazge M., Yusufi M., Devasthale G., and Khan E. (2014) *In vitro* investigation of antibacterial activity of novel 3-acetylcoumarin schiff bases and their molecular docking studies. *Int. J. Pharm. Res. Scholars.*, 3 (1), 696-703.

